

Prevalence and Diversity of Internal Cestode Parasites Infected Nile Tilapia (*Oreochromis niloticus*) and African Catfish (*Clarias gariepinus*) in Farmers Fresh Water Ponds in Kenya

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Abstract

This study is one of series of paper expected to occur in this journal (Part of my PhD work Moi University 2014-17). The study conducted between January 2016--June 2016 involved three counts, Kirinyaga, Kisii, and Uasingishu of Kenya, with the aim to investigate problems of stunted and mortality of 520 fishes of these two species of fresh water from farmer ponds were randomly sampled and examined for internal cestode parasites. 100 fish were *C. gariepinus* and 420 *O. niloticus*. These specimens were killed by scissors or a knife and then dissected internal organs placed in Petri dish mixed well with sodium chloride solution different parasites observed and identified using common characteristics and standard keys. The overall results show that 240 (46.2%) of the total fish examined were infected by different parasites. *C. gariepinus* had a higher infection rate 55 (55.0%) out of the 100 fish sampled compared to *O. niloticus* 185 (44.1%) of the 420 fish sampled. Prevalence and diversity of internal parasites. *Diphyllbothrium latum* (34.59%), *Proteocephalus* spp (49.62%), *Caryophyllidea* spp (13.53%) and mixed infection (2.26%) Statistically Significance was tested at 0.05, The study findings show that there was a significant relationship ($p=0.0002$) between the type of fish and the number of parasites. Also insignificant Relationship observed between internal cestode parasites and sampled fish on length (0.06657), weight (0.1690) and sexes (0.3668). The study concluded that the *C. gariepinus* are more attacked by internal cestode parasites compared *O. niloticus*. Our study suggest that farmers to keep more species with higher tolerance with parasites.

Key words: Internal cestode parasites; stunted and mortality.

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1. Introduction

The assessment of the prevalence of internal cestodes parasites on Nile tilapia (*Oreochromis niloticus*) and African catfish (*Clarias gariepinus*) is very important to the understanding of the functional role and effect of parasites on the two warm water fish species within their living environment [1].

Fish serves as an important source of energy, protein and a range of essential nutrients, accounting for about 17% of the global population's intake of animal protein [1; 16]. In the natural situation where the fish live only few reach adult stages, while majority are predated upon or die due to parasitic infections and other diseases [2]. Further studies by [14] indicated that many fish die under culture conditions than would in the wild due to stress caused by lack of sufficient oxygen, predators and parasitic infection [12; 20]. Low oxygen levels, increased organic matter load and poor aquatic environmental conditions are conditions known to enhance parasitic infections in aquatic systems [2; 19].

Despite the fact that fish in their natural environment experience parasitic infections just like other animals, the presence of fish parasites plays a major role on the fisheries and the fishing industry [7; 11; 32]. There is literature on diagnosis of epizootic infection in both cold and warm water fish species [7; 11; 29]. Even though there are few reported cases of epizootic fish parasites, parasitized fish are more likely to experience mass mortality, retarded growth and weight loss and eventually low market value [2]. Despite fish being a good source of animal protein, fish parasitic infections pose a major threat to aquaculture sub-sector that has recently demonstrated tremendous growth in fish production in mitigating dwindling stocks of global capture fisheries [22; 34]. Studies have indicated that environmental factors have a major impact on fresh water biota and thus play a major role in determining the health of fish [20; 32].

Fish parasites, specifically cestodes are known to inhibit nutrient absorption and is more pronounced in cultured species due to overstocking coupled with poor water quality management [14; 15]. Parasites are important components of any ecosystem and play key roles in fish population dynamics, community structure, and also provide important information on environmental stress, food web structure, function and biodiversity [31; 33]. Parasites take a significant proportion of list of fish diseases (80%) and are among the major causes of economic losses in the fish industry and in aquaculture [14; 15].

Kenya's fish farming has experienced great growth since 2009 as a result of the Economic Stimulus Program [10]. Increasing fish production without improving management of the fish ponds may result in bacterial, viral, fungal and parasitic infestations. On average 80% of fish diseases are parasitic especially in warm water fish, and severe fish mortalities as a result of parasites have been reported elsewhere [5; 7] but few studies have been carried out in Africa [15; 23]. The effect of parasites on the economic value of fish is greater than their impact on human health [35]. Parasites affect the fish host through destroying tissue, removing blood and cellular fluids, diverting part of its nutrient supply and allowing secondary infections to develop easily [4; 24]. Factors that enhance parasitic infection include nature of ponds, quality of feed, culture systems, and low oxygen levels resulting from poor pond management [2]. This knowledge will help fish growers improve their productivity, fish and fish product value, enhance incomes and thus alleviate poverty and malnutrition

Objective of study

To assess the diversity and prevalence of internal parasites in *O. niloticus* and *C. gariepinus* under different culture systems

Research Hypothesis

H₀ There is no significant difference on the diversity and prevalence of internal cestodes parasites in *O. niloticus* and *C. gariepinus* under different culture systems in fresh water ponds.

Significance of the study

The findings in the current study have provided empirical data and will address the challenges posed by internal cestodes parasites of *O. niloticus* and *C. gariepinus* which are important to fish farmers, fisheries and policy makers

2. Materials and methods

Study Area

This study was conducted in three counties in Kenya: Kirinyaga, Kisii and UasinGishu. Kirinyaga County is located in the central region of Kenya approximately 117km Northeast of Nairobi. It lies at latitude N0° S37° and an altitude of 1030 metres above sea level [21; 25]

UasinGishu County is located in the north rift region of Kenya. It lies at latitude N1° E34° and an altitude of 2000 meter above sea level.

Kisii County is located in Nyanza region 64 km from Kisumu city. It lies at latitude 0.6773° S and Longitude 34.7796° E, at an altitude of 2000 meters above sea level.

Study design and sampling method

The study conducted cross sectional and experimental for data collection, Sampling directed by financial and time constraints. Purposive used i.e. calculated samples randomly taken with minimum error hence can be generalised.

520 live fish comprising of (Kirinyaga=144*O. niloticus* and 40 *C. gariepinus*), (Kisii= 162*O. niloticus* and 30 *C. gariepinus*) and (UasinGishu= 114*O. niloticus* and 30 *C. gariepinus*) were sampled for parasites analysis. The box like containers (1Mx1Mx1M) half filled with water was used for transported alive fish sampled from field to laboratory for parasites examination. The sample size for this field study was calculated using the formula by [25].

Measurement and morphological description of fish

The study involved morphological description of fish means used data obtained from body of fish samples (sexes, length, weight and maturity).

The weights of all sampled fish were measured using an electronic balance TX4202L and their length measured using a meter ruler and recorded. The sex of sampled fish was determined by observing the genital organs and the observation was recorded for each fish specimen

.Maturity was determined by pressing the abdomen for sperms for adult male fish and eggs, sometimes spawned (young) in mouth for adult female fish .Absence of sperms or eggs were an indication that the fish were immature.

The sex of the fish was also determined by observing the testes and ovaries (gonad) after dissecting the specimen. [14; 36]

Parasitological examination (gastrointestinal parasites)

Fish specimens were dissected from the anus to the lower jaw using a pair of dissecting scissors. All visceral organs i.e., stomach, intestines and the liver were extracted and placed on a Petri dish and bathed with sodium chloride solution 9 gms/l for parasites recovery.

The stomach and entire intestine were dissected longitudinally with a pair of dissecting scissors and then placed in sample bottle which was shaken vigorously for about 1-2 minutes thereafter the upper part of the mixture was discarded. This process was repeated until the remaining liquid was transparent the parasites that were visible to the naked eye were placed on a glass slide and transferred to the SZ-ST Olympus stage microscope for examination and identification. Photos of the observed parasites were taken using an Olympus camera PM-6 230154 mounted on the dissecting microscope. The parasites that were identified were recorded. The parasites were identified by observing common characteristics and by the support of standard keys [31;36;37] The parasites were then placed in labeled sample bottles (vials) containing 70% alcohol for preservation.

Data analysis

The data of this study of all counts collected from field were coded and stored in Microsoft excel. Finally these data were analyzed using MINITAB- Statistic software, 14 Statistical Significance was tested at 0.05, Fisher Exact test were used to test for significance between length, weight, and sexes.

3. Results

A total of 520 live fish comprising of 420 *O. niloticus* and 100 *C. gariepinus* were sampled and examined for internal cestodes parasites. 144 *O. niloticus* and 40 *C. gariepinus* were sampled in Kirinyaga, 162 *O. niloticus* and 30 *C. gariepinus* in Kisii while in UasinGishu, a total of 114 *O. niloticus* and 30 *C. gariepinus* were sampled and examined for internal cestodes (Figure 1)

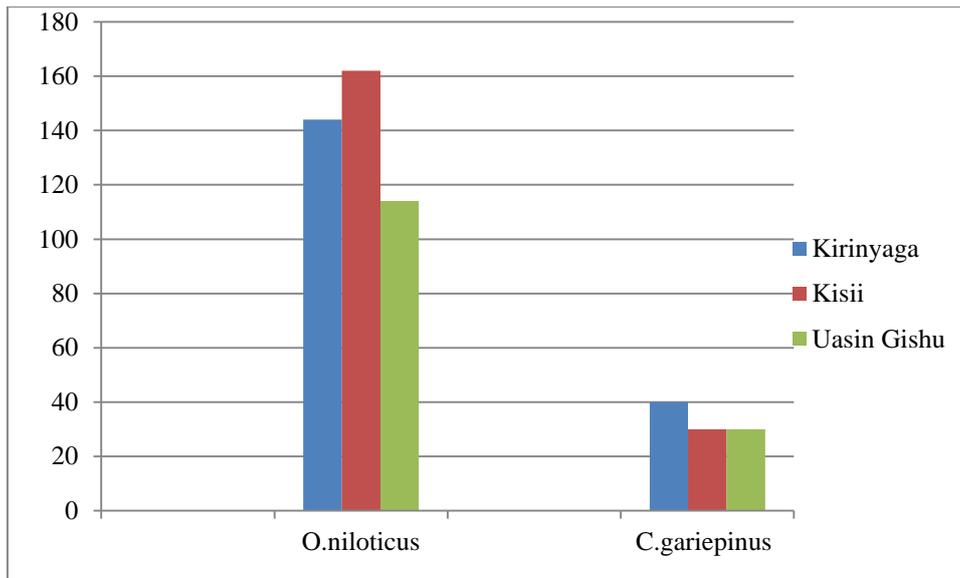


Figure1: showing fish species and total number of fish sampled per County.

Table 1: Diversity and prevalence of internal cestode parasites in *O. niloticus* and *C. gariepinus*

Fish species	None infected	Internal cestodes Parasites			Total infected
		Between 1 and 4	Between 5 and 8	Between 9 and 12	
<i>O. niloticus</i>	235 56.0%	173 41.2%	12 2.9%	0 0.0%	185 44.1%
<i>C. gariepinus</i>	45 45.0%	42 42.0%	10 10.0%	3 3.0%	55 55.0%
Total	280 53.8%	215 41.3%	22 4.2%	3 0.6%	240 46.2%

The results show that 240 (46.2%) of the total fish examined were infected by different parasites. *C. gariepinus* had a higher infection rate 55(55.0%) out of the 100 fish sampled compared to *O. niloticus* 185 (44.1%) of the 420 fish sampled.

The results in table 2 indicated a significant difference ($p = 0.0002$) for Fisher Exact test between the type of fish and the number of parasites in the fish. In this case *C. gariepinus* had more likelihood of attacks by parasites compare to *O. niloticus*

Table 2: Statistical relationship showing diversity and prevalence of internal cestode parasites between *O. niloticus* and *C. gariepinus*.

Tests for association of variables: Fisher and Chi-Square Tests			
	Value	Df	statistical
Fisher Exact test	-	-	0.0002
Chi-square test:			
Likelihood Ratio	19.567	15	0.1630
Linear-by-Linear Association	12.684	11	0.9230
N of Valid Cases	520		

Table 3: Diversity and prevalence of internal cestodes parasites in examined fish species

Fish species	Caryophyllaeidea species	Cestodes			Total
		Proteocephalus species	Diphyllobothrium species	2 Cestodes	
<i>O. niloticus</i>	15 13.8%	55 50.5%	37 33.9%	2 1.8%	109 100%
<i>C. gariepinus</i>	3 12.5%	11 45.8%	9 37.5%	1 4.2%	24 100%
Total	18 13.5%	66 49.6%	46 34.6%	3 2.3%	133 100%

Table 3: indicates that Caryophyllaeidea species was only found in *O. niloticus*. The study also indicated that, the Proteocephalus species was highly distributed in *C. gariepinus* compared to *O. niloticus* while the distribution of Diphyllobothrium species was almost the same in both *O. niloticus* and *C. gariepinus* i.e., 33.90% and 34.60% respectively.

The results show that. Infection was higher in immature fish (49.0%) compared to mature fish (45.4%) as shown in table 4.

The results of this finding indicated that internal cestodes parasites were more prevalent in fish of length 21cm and below compared to those of length ≥ 22 cm. Generally infection decreases with increasing length across all internal cestodes species as shown in table.6.

Table 4: Distribution of internal cestode parasites by maturity of the fish examined.

Infection Status	Maturity		Total
	Mature	Immature	
None	227	53	280
	81.1%	18.9%	100.0%
Infected	189	51	240
	45.4%	49.0%	46.2%
Total	416	104	520
	80.0%	20.0%	100.0%

Table 5: Diversity and prevalence of internal cestode parasites in relation to length of fish examined.

Internal cestode	Length of Fish					Total
	< 14cm	14cm≤21cm	22≤28cm	29≤35cm	>35cm	
Caryophyllaeidea species	3	6	6	3	0	18
	16.70%	33.3%	33.3%	16.0%	0.0%	100%
Proteocephalus species	8	43	7	7	1	67
	12.1%	63.6%	10.6%	10.6%	1.5%	100%
Diphyllobothrium species	5	30	3	8	0	46
	10.9%	65.2%	6.5%	17.4%	0.0%	100%
2 Cestodes	0	2	0	0	1	3
	0.0%	66.7%	0.0%	0.0%	33.3%	100%
Total	16	81	16	18	2	133
	12.0%	60.2%	12.0%	13.5%	1.5%	100%

Table 6: Statistics showing diversity and prevalence of internal cestode parasites in relation to length of fish examined.

Tests for association of variables: Fisher and Chi-Square Tests			
	Value	df	statistical
Fisher Exact test			0.06657
Chi-square test:			
Likelihood Ratio	+20.23	15	0.163
Linear-by-Linear Association	0.009	1	0.923
N of Valid Cases	520	-	-

The results indicated that there was no significant relationship ($p = 0.06657$) between the length of fish and diversity and prevalence of internal cestode parasites. Although it was observed that infection with internal cestode parasites tailed -off with increasing length (table.6), this was not statistically significant as shown in table8.

Table 7: Distribution and prevalence of internal cestode parasites in relation to weight of fish examined.

Parasites Counts	Fish Weight (grams)						Total
	<50	50≤100	101≤150	151≤200	201≤250	≥250	
None	48 17.1%	98 35.0%	71 25.4%	35 12.5%	21 7.5%	7 2.5%	280 100%
Between 1 and 4	26 12.1%	79 36.7%	48 22.3%	31 14.4%	24 11.2%	7 3.3%	215 100%
Between 5 and 8	0 0.0%	5 22.7%	3 13.6%	7 31.8%	6 27.3%	1 4.5%	22 100%
Between 9 and 12	0 0.0%	0 0.0%	0 0.0%	3 100.0%	0 0.0%	0 0.0%	3 100%
Total	74 14.2%	182 35.0%	122 23.5%	76 14.6%	51 9.8%	15 2.9%	520 100%

The results of this finding indicated that internal cestodes parasites were more prevalent in fish of weight 200grams and below compared to those of weight ≥ 201 grams. Generally infection decreases with increasing weight across all internal cestodes species examined. It was also observed that fish that weight less than 50grams only harbor between 1 and 4 internal cestodes parasites. The occurrence of between 9 and 12 internal parasites was only recorded in fish that weight between 151 and 200 grams as shown in table

Table 8: Statistics showing distribution and prevalence of internal cestode parasites in relation to weight of fish examined.

Tests for association of variables: Fisher and Chi-Square Tests				
	Value	df	statistical	sign(p=)
Fisher Exact test	-	-	0.169	
Chi square test				
Likelihood Ratio	34.37	15	0.163	
Linear-by-Linear Association	14.134	1	0.923	
N of Valid Cases	520	-	-	

The results indicated that prevalence and diversity of internal cestode parasites in relation to weight of fish do not differ significantly ($p = 0.196$). Although fish of weight 200grams and below recorded more infections compared to those of weight ≥ 201 grams this was not statistically significant. Generally infection decreases with increasing weight across all internal cestodes species examined as shown in table .

Table 9: Sex-related diversity and prevalence of internal cestode parasites species in examined fish.

internal cestodes	Sex		Total
	Male	Female	
Caryophyllaeidea	15	3	18
species	83.3%	16.7%	100%
Proteocephalus species	49	17	66
	74.2%	25.8%	100%
Diphyllobothriumlatum	27	19	46
species	58.7%	41.3%	100%
2 Cestodes	2	1	3
	66.7%	33.3%	100%
Total	93	40	133
	69.9%	30.1%	100%

The study results indicated that Caryophyllaeidea species occur most frequently in males (83.30%) with the least frequently occurring internal cestodes parasites to be Diphyllobothriumlatum (58.7%). Whereas in females, Diphyllobothriumlatum (41.30%) had the highest prevalence with the lowest by Caryophyllaeidea species (16.7%) as shown in table 9

Table 10: Sex-related distribution and prevalence of internal cestode parasites in examined fish.

Tests for association of variables: Fisher and Chi-Square Tests				
	Value	df	Statistical Sign (p=)	sign(p=) .
Fisher Exact test	-	-	0.3668	
Chi square test:				
Likelihood Ratio	12.286	15	0.056	
Linear-by-Linear Association	1.996	1	0.158	
N of Valid Cases	520	-	-	-

The study findings indicated that there was no significant relationship between the prevalence and diversity of internal cestode parasites and the sex of fish with Fisher Exact test ($p=0.3668$) and Linear by Linear Association ($p=0.158$) as shown in table 10. Even though female fish harbor more diverse parasites than the male, this was not statistically significant.

4. Discussion

The findings indicated that 240 (46.2%) of the total fish examined were infected by different internal cestodes parasites. These are *Proteocephalus species*, *Diphyllbothriumlatum*, and *Caryophyllaeidea species*. This percentage prevalence is consistent with studies conducted by (38;8) in fresh water ponds in Lahore Pakistan in which they reported a prevalence of 44.45% infection by internal cestodes parasites in nine (9) fish samples. In India, (8) have reported a slightly lower prevalence of internal cestodes parasites of between 26.31% and 36.84% at different sampling sites during the monsoon season. Similarly (3) have reported a prevalence of gastrointestinal parasites infection of 38.7% in Nigeria. They attributed these to poor hygienic conditions of the ponds which tend to favour the existence and propagation of many parasites. Decaying of the organic matter in the bottom of the ponds, depletes dissolved oxygen and water quality parameters are altered. The too could be the case in the present study where most of the ponds used were not properly maintained. Related studies conducted by (40) in Maiduguri, Nigeria have recorded similar parasites prevalence of 38.6% in *C. gariepinus* though fish sampled were purchased from local fish sellers within the metropolis. In Kenya (25), carrying out studies at KMFRI Sagana Aquaculture Research Centre and river Tana and, (17) in Lake Baringo have reported higher percentage prevalence (65.8% and 68.33%) of helminthes parasites. Helminthes infestation at a prevalence of 77.7% and 60.3% in Gandoman Lagoon, Iran and river Niger of Nigeria respectively have been reported (29;16). This high cestode prevalence could be attributed to the fact that these studies were carried out in rivers, lake and lagoon which are apparently dirty and have free materials deposits caused by eroded substances that may make the environments conducive for parasite development. The difference could also be that these studies recorded all helminthes of internal parasites whereas in the current study, only internal cestodes parasites were reported. The current study showed that *C. gariepinus* had the highest prevalence and diversity of worm burdens (internal cestodes parasites) of 55(55.0%, n=55) compared to *O. niloticus* 185 (44.1%, n=185). This difference could be attributed to the habitat favoured by *C. gariepinus* which consists of turbid environments and shore areas that are usually covered with vegetation (27), as was the case at many of the ponds used for this study. These habitats favour the intermediate hosts of cestodes as well as other digenetic trematodes (40). Further, *C. gariepinus* are omnivorous fish that feed on both aquatic plants and animals including copepods (28). Copepods act as first intermediate host of most internal parasites that infest freshwater fish (26;30). The larval stage of internal cestode parasites gains entry into copepods through ingestion as the copepod seeks food within its habitat. Copepods carrying the larval stage of cestodes are thereafter fed on by fish which act as second or final host that complete the lifecycle of cestodes (17). The large size of *C. gariepinus* also predisposes this particular species to parasites infestation compared to those of *O. niloticus* (27). The presence of fewer internal cestodes parasites in *O. niloticus* could be attributed to its resistance to parasitic infections. This may be explained by the fact that *O. niloticus* species rarely succumb to disease epidemics and have a remarkable power of recovery from infections (35). Therefore this could account for the low infection rates recorded during this study regardless of poor management to some of the ponds. These cestodes are widely reported around the world. For instance *Proteocephalus*, and *Caryophyllaeidea species* have also been reported in India (7) and Kenya (25) in fish sampled from rivers, lakes and fish ponds. Studies conducted in Zimbabwe by (27) have reported *Polyonchobothriumclarias* and *Proteocephalusglanduliger*. The tapeworm species (*Diphyllbothriumlatum*) reported in this study also had recorded current study in Africa.

Diphyllobothrium latum was found in *C. gariepinus* in Maiduguri, Nigeria (40). The possible reasons for the recovery of these fish tapeworms in Africa of recent is that most small fish farmers use rivers water in their ponds, this water may carry first intermediate host copepods hence circulate from country to country. Another possible reason is distribution through the intermediate host (definitive or final host) of cestode e.g. piscivorous bird, mammals or reptiles (17). Researcher of present study recommended that further molecular and phylogenetic be conducted to confirm *Diphyllobothrium latum* species reported. This could be that more researches are being done in the areas of fish parasites of recent in Africa. The findings in relation to fish maturity clearly show that infection was higher in immature fish (49.0%, n=51) compared to mature fish (45.4%, n=189) in *O. niloticus* and *C. gariepinus* in all the three (3) counties of Kirinyaga, UasinGishu and Kisii. The possible reason for this relationship immature fed less amount of foods hence gained less immunity compared the mature fish. This is in agreement with study conducted in Nigeria (3) who reported that smaller (immature) fish were more infected compared to larger (mature) probably due to their nature of acquired immunity with age. In contrast, the present study disagrees with findings reported (41;16) who both reported that bigger (and therefore possibly mature) fish have more parasites compared to small fish because they feed more on diverse food sources thereby exposing them to more parasitic infestation. The present study findings in relation to fish length indicated that internal cestodes parasites were more prevalent in fish of length 21cm and below compared to those of length ≥ 22 cm. Generally infection decreases with increasing length of fish to all types of internal cestodes parasites investigated. Internal cestodes parasites were more prevalent in fish of weight 200grams and below compared to those of weight ≥ 201 grams. The results obtained in the current study is in agreement with those reported (42) They reported that infestation was lowest in fish with body length of between 23.00 to 25.75 cm and highest in fish with 25.76 to 31.25 cm body length. Similarly, the parasitic infestation increased with lower body weight of between 160-258 g, while almost no parasites were observed in heavier fish (>553 g). This may be attributed to the fact that fish of a shorter length and lower weight, and therefore possibly immature may not have acquired immunity compared to fish with large length and heavier weight fish (mature). Larger fish, in the current study which had experienced exposure to different internal parasites might have acquired stronger immunity hence their higher capacity to resist infestation of parasites (42). Contrary to the present reports, (3) in Nigeria and (30) in Egypt have both reported that the smaller fish are relatively less infected than larger ones. This they also attributed to long time accumulation of parasites by larger and matured fish compared to the smaller/young ones. In relation to the sex of the fish, more males were infected with internal cestodes parasites (68.3%) compared to the female fish (31.5%) in both *O. niloticus* and *C. gariepinus*. However no variation or significant different in prevalence and diversity of cestodes was observed between the sexes. A high rate of infection was observed in males which may be attributed to activeness and aggressiveness of males compared to female fish which gives them an upper hand in the search for food hence increasing their chance of acquiring more internal cestodes parasites compared to female fish. Similar studies by Omeji and his colleagues (2013) had also reported that male fish from earthen pond had a higher parasitic infestation (64.29%) than the female fish (57.69%). Similarly, (3) also recorded higher infection levels in male fish (37.8%) than female fish (23.5%). On the other hand, in Female fish cultured in concrete ponds in Nigeria were reported to have a higher percentage of parasitic infestation (22.73%) than the male (16.67%) (28). Abdel-gaber and his colleagues (2015) in Egypt also recorded higher prevalence rate 72 (90%) in female fish than males 58 (48.33%) although the difference was not significant ($P > 0.05$). This was attributed to the

physiological state of the female fish, as most gravid females fish could have had reduced resistance to infestation by parasites. In addition, their increased rate of food intake to meet their food requirements for the development of their eggs might have exposed them to more contact with the parasites, which subsequently could increased their chance of being infected

5. Conclusion and recommendation

The results of this study recorded high incidence of infections of the internal cestode parasites prevalence and diversity in both species sampled *O. niloticus* and *C. gariepinus*. Therefore stake holders should train the farmers effect of these parasites before starting keeping fish. The researcher of this study suggests more studies on internal parasites to be conducted

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Reference

- [1] Abila, O. R. (2003). Case study: Kenya fish exports. In: Food Safety in Food security and Food Trade, International Food Policy Research Institute, Washington D. C
- [2] Abowei, J. F. N., Briyai, O. F., & Bassey, S. E. (2011). A Review of Some Basic Parasite Diseases in Culture Fisheries Flagellids, Dinoflagellides and Ichthyophthiasis, Ichtyobodiasis, Coccidiosis Trichodiniasis, Heminthiasis, Hirudinea Infestation, Crustacean Parsite and Ciliates, 2(5), 213–226.
- [3] Akinsanya 1 B, A.A. Hassan2 & A.O. Adeogun (2008) Gastrointestinal Helminth Parasites of the fish *Synodontis Clarias* (Siluriformes: Mochokidae) from Lekki lagoon, Lagos, Nigeria [4] Akoll, P., Konecny, R., Mwanja, W. W., Nattabi, J. K., Agoe, C., & Schiemer, F. (2011). Parasite fauna of farmed Nile tilapia (*Oreochromis niloticus*) and African catfish (*Clarias gariepinus*) in Uganda. <http://doi.org/10.1007/s00436-011-2491-4>.
- [5] Amare A, Alemayehu A, & Aylate A (2014) Prevalence of Internal Parasitic Helminthes Infected *Oreochromis niloticus* (Nile Tilapia), & *Clarias gariepinus* (African catfish)
- [6] Bellay, S., Oliveira, E. F. D. E., & Mello, M. A. R. (2012a). Ectoparasites and endoparasites of fish form networks with different structures. Bellay, S., Oliveira, E. F. D. E., & Mello, M. A. R. (2012). Ectoparasites and endoparasites of fish form networks with different structures. <http://doi.org/10.1017/S0031182015000128>
- [6] Bellay, S., Oliveira, E. F. D. E., & Mello, M. A. R. (2012b). Ectoparasites and endoparasites of fish

- form networks with different structures. <http://doi.org/10.1017/S0031182015000128>
- [8] BhureDhanrajBalbhim and Sanjay ShamraoNanware(2014) Studies on prevalence of cestode parasites offreshwater fish, Channapunctatus. Journal of Entomology and Zoology Studies 2014; 2 (4): 283-285
- [9] Bob-manuel, F. G., & Harcourt, P. (2013). Food and Feeding Habits of Tilapia guineensis(1862) in Rumuolumeni Creek , Niger Delta : Implications for Pisciculture, 5(1), 41–45
- [10] Charo-Karisa H and Gichuri M. (2010). Overview of the Fish Farming Enterprise Productivity Program. In: End of Year Report Fish Farming Enterprise Productivity Program Phase I. Aquaculture Development Working Group Ministry of Fisheries Development, Nairobi, KE
- [11] Choundhury, A .,Charipar, E., Nelson, P .,Hodgson, J. R., Bonar, S. and Cole,R. A. (2006). Update on the distribution of the invasive Asian fish tapeworm, Bothriocephalusacheilognathi, in the U.S. and Canada. ComparativeParasitology, 73: 269–273. [5]
- [12] Corps, P. (1976). Freshwater Fish Pond Culture & Management
- [13] Doreen Z. Moyo, ChakanetsaChimbira and PhumuzileYalala(2009) Observations on the Helminthes Parasites of Fish in Insukamini Dam, Zimbabwe Research Journal of Agriculture and Biological Sciences, 5(5): 782-785
- [14] Edema, C. U., Okaka, C. E., Oboh, I. P., & Okogub, B. O. (2008). A preliminary study of parasitic infections of some fishes from Okhuo River , Benin City , Nigeria ., 4(3), 107–112.
- [15] El-seify, M. A., Zaki, M. S., Desouky, A. R. Y., Abbas, H. H., Hady, O. K. A., & Zaid, A. A. A. (2011). Seasonal Variations and Prevalence of Some External Parasites Affecting Freshwater Fishes Reared at Upper Egypt, 8(3), 397–400.
- [16] Eyo Joseph Effiong and Florence OyiboIyaji (2014) Parasites of clarotesLatticeps (Reppull 1832 Siluriforms, Bagridie) at River Niger-Benue Confluence lokoje, Nigeria. Journal of Fisheries and AquatScience 9(3) 125-133.
- [17] .Gumpinger, (2016) Parasite Species Richness of fish from Lake Baringo, Kenya
- [18] Hine PM (1975). Final report on investigation into diseases and parasites of wild and farmed eels in South Africa. Report to JLB Smits Institute of Ichthyology, Grahams town, Republic of South Africa. Jackson PBN (1978). Health and disease in intensive aquaculture. J.S.A. Vet. Ass., 49: 57–59.
- [19] Horwitz, P., and Wilcox, B. A. (2005). Parasites, ecosystems and sustainability: An ecological and complex systems perspective. International Journal for Parasitology, 35: 725–732
- [20] Justine, J., Briand, M., Bray, R. A., Justine, J., Briand, M., &A, R. A. B. (2012). A quick and simple

method , usable in the field , for collecting parasites in suitable condition for both morphological and molecular studies.

- [21] Jacob, J. H., Anger, J. Vitale., Srinarasan,(2007) Economic damage in Kenyas upper of Arc ViewSweat.Journal of spatial Hydrology.7:1.
- [22] Khalil, M. I, El-Shahawy, I. S, and Abdelkader, H. S (2014). Studies on some fish parasites of public health importance in the southern area of Saudi Arabia. *Braz. J. Vet. Parasitol.*, Jaboticabal, v. 23, n. 4, p. 435-442.
- [23] León, G. P. De, Rosas-valdez, R., Aguilar-aguilar, R., Mendoza-, B., Pérez-rodríguez, R., & Domínguez-domínguez, O. (2010). Helminth parasites of freshwater fishes , Nazas River basin , northern Mexico.
- [24] Lugo, L. (2014). Prevalence of Internal Parasitic Helminthes Infected *Oreochromis niloticus* (Nile Tilapia), *Clarias gariepinus* (African Catfish) and *Cyprinus*, 5(3). <http://doi.org/10.4172/2155-9546.1000233>
- [25] Mathenge. (2010). Prevalence , Intensity And Pathological Lesions Associated With Helminth Infections In Farmed And Wild Fish In Upper Tana River Basin , Kenya Charles Gichohi Mathenge (Bvm , Uon).
- [26] Mdegela **R.H** , OmaryA N, Mathew.C and NongaH E,(2011) Effect of Pond Management on Prevalence of Intestinal Parasites in Nile Tilapia (*Oreochromis niloticus*) under Small Scale Fish Farming Systems I Morogoro, Tanzania [Livestock Research for Rural Development. 23 \(6\) 2011](#)
- [27] Moyo Doreen Z. , ChakanetsaChimbira and PhumuzileYalala(2009) Observations on the Helminthes Parasites of Fish in Insukamini Dam, Zimbabwe. *Research Journal of Agriculture and Biological Sciences*, 5(5): 782-785
- [28] Omeji,S.,S.G.SolomonAndUloko,C(2013)Comparative Study On The Endo-Parasitic Infestation In *ClariasGariepinus* Collected From Earthen And Concrete Ponds InMakurdi, Benue State, Nigeria *IOSR Journal of Agriculture and Veterinary Science (IOSR-JAVS)*ISSN: 2319-2380, ISBN: 2319-2372. Volume 2, Issue 1 PP 45-49www.iosrjournals.org
- [29] Raissy, M., Ansari, M., Lashkari, A., & Jalali, B. (2010). Occurrence of parasites in selected fish species in Gandoman Lagoon , Iran, 9(3), 464–471.
- [30] Rewaida Abdel-Gaber*, Manal El Garhy,& Kareem Morsy(2015) Prevalence and Intensity of HelminthParasites of African Catfish *Clariasgariepinus* in Lake Manzala, Egypt *Advances in Bioscience and Biotechnology*,2015, 6, 464-469
- [31] Scholz, T., . Kuchta, R., A Brabec, J., and Bray, R. (2011). Suppression of the tapeworm

- order Pseudophyllidea (Platy helminthes: Eucestoda) and the proposal of two new orders, Bothriocephalidea and Diphyllbothriidea. *International Journal for Parasitology*, 38: 49–5
- [32] Sriyasa, P., Chitmanat, C., Whangchai, N., Promya, J., Lebel, L., & Abbas, H. H. (2013). Effects of Temperature upon Water Turnover in Fish Ponds in Northern Thailand, 2013 (September), 18–23.
- [33] Waeschenbach, A., Webster, B. L., and Littlewoods, D. T. J. (2012). Adding resolution to ordinal level relationships of tapeworms (Platy helminthes: Cestoda) with large fragments of mt DNA. *Molecular*
- [34] FAO, 2015. FAO Global Aquaculture Production database updated to 2013 – 2014 Summary information.
- [7] Ash, A. (2012). Diversity of tapeworms (Cestoda) in freshwater fish of India. Ph.D. Thesis, in English, Faculty of Science, University of South Bohemia in České Budějovice, Czech Republic
- [35] Ayanda, O. I. (2009). Comparative parasitic helminth infection between cultured and wild species of *Clarias gariepinus* in Ilorin, North – Central Nigeria. *Scientific Research and Essay*, 4: pp. 018-021, ISSN 1992-2248. Accessed at <http://www.academicjournal.org/SRE> on 2nd May 2009, 176 pp
- [36] Bush A. O., Lafferty K. D., Lotz J. M., Shostak A. W., 1997, Parasitology meet ecology on its own terms: Margolis et al. revisited, *J. Parasitology.*, 83 (4), 575–583.
- [37] Yagamuti, 1934 in yellow fish in the Vaal Dam, South Africa. *Onderstepoort Journal of Veterinary Research*, 72, 207–217.
- [38] Muhammad Shafiq Ahmed, Tahir Iqbal Anjad, Mahmood Muhammad Gulzarin and Muhammad Abid, (2007) helminth parasites of some freshwater fishes Punjab Univ. *J. Zool.*, Vol. 22 (1-2), pp. 01-06, 2007
- [40] Biu, A. A., & Akorede, G. J. (2013): Prevalence of Endoparasites of *Clarias gariepinus* (Burchell 1822) in Maiduguri, Nigeria. *Nigerian Journal of Fisheries and Aquaculture*, 1:1–6
- [41] Yimer, E. (2000). Preliminary survey of Parasites and Bacterial Pathogens of fish at Lake Ziway. *Ethiopian Journal of Science*, 23(1): 25–33 [42] Tadesse, B. (2009): Prevalence and abundance of fish parasites in Bomosa cage systems and Lakes Babogaya and Awasse, Ethiopia. Thesis (M. Sc.): UNESCO-IHE, The Netherlands
- [43] Ashade OO, Osineye OM, Kumoye EA (2013). Isolation, Identification, and Prevalence of Parasites on *Oreochromis niloticus* from three selected River Systems. *Journal of Fisheries and Aquatic Science*, 1(8): 115-121.